Derivatives of 2-[[N-(Aminocarbonyl)-N-hydroxyamino]methyl]-1,4-benzodioxan as Orally Active 5-Lipoxygenase Inhibitors

Yoshitaka Satoh,* Colleen Powers, Leticia M. Toledo, Timothy J. Kowalski, Paul A. Peters, and Earl F. Kimble Research Department, Pharmaceuticals Division, CIBA-GEIGY Corporation, 556 Morris Avenue, Summit, New Jersey 07901

Received August 15, 1994[®]

N-Hydroxyureas based on the 1,4-benzodioxan template were prepared from appropriately substituted 1,4-benzodioxan-2-methanols as the key intermediates and evaluated in the in vitro guinea pig polymorphonuclear leukocyte 5-lipoxygenase (5-LO) assay for their 5-LO inhibitory activity. Placement of a 7-phenoxy or 7-p-fluorophenoxy substituent resulted in a dramatic increase in *in vitro* potency. Selected compounds were subsequently assayed in an *ex vivo* dog model of LTB₄ synthesis at a dose of 1.0 mg/kg. The 7-phenoxy derivatives 16 and 17 showed modest duration of action (DA) in this dog model. The 6-regioisomers 21 and 22 were less potent. Replacement of the 7-phenoxy group of 16 with the p-fluorophenoxy moiety enhanced the DA dramatically. Compound 18 (CGS 25667), which had an IC_{50} value of 100 nM in the in vitro guinea pig 5-LO assay, had a DA of 8.5 h (zileuton, DA = 8.5 h) at the oral dose of 1.0 mg/kg. Optical antipodes (24, 26) of 18 were independently synthesized in high (>95%) enantiomeric purity from commercially available optically active glycidyl tosylates and evaluated. In the *in vitro* assay, the 2S-(-)-enantiomer (24, CGS 25997, IC₅₀ = 85 nM) was found to be twice as active as the 2R-(+)-counterpart (26, CGS 25998, IC₅₀ = 180 nM). In the ex vivo experiment, 24, which dose dependently inhibited plasma 5-LO activity, was shown to be significantly longer acting than 26, with a DA of 8.4 h when dosed orally at 1.0 mg/kg.

Introduction

It is becoming increasingly evident that leukotrienes (LT's) play extremely important roles as the major mediators of inflammatory reactions and hypersensitivity in mammalian animals, including human.¹ In vivo, LT's are produced from arachidonic acid (AA) by a series of enzymatic transformations. Since the oxygenation of AA by 5-lipoxygenase (5-LO) is the first committed step for the production of LT's, it has been long assumed that inhibition of 5-lipoxygenase would provide a novel therapy for human inflammatory disease conditions² such as asthma, rheumatoid arthritis, and inflammatory bowel disease. As a result of intense drug discovery efforts in the last decade, several inhibitors of 5-LO, such as zileuton,³ MK-886,⁴ MK-591,⁵ Wy-50,295,⁶ ZD2138.⁷ and so forth, have been identified and entered clinical evaluation (Figure 1). These results indicate promising therapeutic benefits of 5-lipoxygenase inhibition in certain inflammatory conditions.^{8,9}

Due to the iron-containing nature of the 5-LO enzyme, inhibitors based on metal chelators have been extensively studied. Among those tested, hydroxamatederived inhibitors¹⁰ have met with considerable success. Conversion of lipophilic hydroxamic acids to their 'reverse' analogs¹¹ or to the *N*-hydroxyurea derivatives,¹² as demonstrated in the case of zileuton,³ was found to enhance the oral efficacy, largely due to increased *in vivo* stability.

In this article, we would like to disclose some of our continuing efforts to identify highly potent and orally available 5-lipoxygenase inhibitors.¹³ A series of N-hydroxyureas based on a benzodioxan template were synthesized and subsequently evaluated *in vitro* and *in vivo*. Due to the chirality of these compounds, the enantiospecificity aspect of the most potent inhibitor was also investigated.

Chemistry

Benzodioxan N-hydroxyureas with no substituents on the aryl ring system were conveniently prepared from commercially available 1,4-benzodioxan-2-methanol (1) (Scheme 1). Thus, the condensation of the bromide with a sodium salt of N,O-bis(*tert*-Boc)hydroxylamine gave a fully protected hydroxylamine precursor (2). Deprotection with TFA followed by condensation with trimethylsilyl isocyanate gave 4. Reaction of 3 with 1 equiv of alkyl isocyanates resulted in the formation of a mixture of the desired N-hydroxyureas (5–7) and N,Obis-carbamoylated derivatives. Compounds 5–7 were, therefore, prepared by first condensing 3 with 2.2 equiv of an appropriate isocyanate followed by monohydrolysis of the O-carbamoyl group with an aqueous base.

Regiocontrolled synthesis of 6-substituted 1,4-benzodioxan-2-methanols was carried out using 5-substituted salicyl aldehydes (8, 19) as the starting materials according to the procedure described by Henning et al.¹⁴ (Scheme 2). Alkylation of 8 and 9 with epibromohydrin and subsequent Baever-Villiger oxidation with m-CPBA yielded the formyloxy derivatives (10, 11). Treatment of 10 and 11 with hydroxide resulted in hydrolysis of the formate and concomitant clean, regioseletive intramolecular epoxide opening to their 1,4-benzodioxan derivatives (12, 13). When MMPP was used in the oxidation step, free phenols instead of their formates were isolated upon workup. Compounds 12 and 13 were converted to the corresponding N-hydroxyureas (16-18) according to the procedure described in Scheme 1. It should be noted that the condensation of the hydroxylamine (14) with methyl isocyanate yielded N-hydroxyurea derivative 17, uncontaminated with the bisadduct, under the same conditions.

The regioisomeric benzodioxans (21, 22) were prepared from the same intermediate (8) (Scheme 3). Thus, 8 was first protected with benzyl bromide and subjected

[®] Abstract published in Advance ACS Abstracts, December 1, 1994.







zileuton



WY-50,295



ZD2138

Figure 1. Representative 5-lipoxygenase inhibitors.

Scheme 1^a



^a Reagents and conditions: (a) Ph₃PBr₂; (b) Boc^tNHOBoc^t, NaH; (c) CF₃COOH; (d) Me₃SiN=C=O, heat; (e) 2.2 equiv of R¹N=C=O, heat; (f) OH⁻.

Scheme 2^a



^a Reagents and conditions: (a) epibromohydrin, K_2CO_3 , DMF; (b) MMPP or *m*-CPBA (see text); (c) OH⁻; (d) MsCl, Et₃N; (e) NaI, MEK; (f) Boc^tNHOBoc^t, NaH; (g) CF₃COOH; (h) Me₃SiN=C=O; (i) CH₃N=C=O.

to Baeyer-Villiger oxidation with hydrogen peroxide. The phenol thus formed was treated with epibromohydrin to give the tris-ether (19). Catalytic hydrogenation of 19 with palladium hydroxide (atmospheric H₂ in ethyl acetate) gave the debenzylated phenol which cyclizes upon treatment with aqueous base to yield 20. It is extremely important to monitor the hydrogenation closely since reductive opening of the epoxide function becomes a significant side reaction under certain conditions.¹⁵ Compound **20** was transformed into the desired *N*-hydroxyureas (**21**, **22**) under the standard conditions described above.

Scheme 3^a



21: R³ = H 22: R³ = CH₃

^a Reagents and conditions: (a) BnBr, K₂CO₃; (b) MMPP; (c) epibromohydrin, K₂CO₃; (d) H₂, Pd/C; (e) OH⁻.

Scheme 4



This method is easily applicable for the synthesis of optically active 2-substituted 1,4-benzodioxans.^{16,17} Thus, commercially available (2R)-(-)-glycidyl tosylate (23) and its 2S-(+)-antipode (25) were converted to the corresponding optically active N-hydroxyureas 24 and **26** (Scheme 4), through the key intermediates 2S- and 2R-13, respectively. The stereochemical consequence was assigned on the basis of well-precedented S_N2 substitution of glycidyl arenesulfonates with phenoxides¹⁸ and the inversion of the stereochemistry upon intramolecular epoxide opening.^{16c} The optical purity of 2S- and 2R-13 was estimated to be 95% by the comparison of their Mosher esters by ¹H NMR.¹⁹ This methodology appears to be highly useful for the synthesis of unsymmetrically substituted, optically active 1,4-benzodioxan-2-methanols with predictable stereochemical outcome.

Biological Results and Discussion

In vitro 5-LO inhibitory activity was determined by measuring production of 5-hydroxyeicosatetraenoic acid (5-HETE) and LTB₄ in guinea pig polymorphonuclear leukocytes (PMN's).¹³ The assay is based on A23187stimulated conversion of [¹⁴C]AA in leukocytes to form the desired products, which are measured by a radiometric thin-layer chromatographic assay. The IC₅₀ values for both products were calculated as the concentration of the test compound at which the synthesis of 5-HETE and LTB₄ were reduced to 50% of their respective control values. The results are summarized in Table 1.

Our previous results in the chromene-based N-hydroxyurea series¹³ indicated either the phenoxy or p-fluorophenoxy substituents on the core substituent

Table 1. In Vitro Guinea Pig PMN 5-LO Assay

	molecular			$IC_{50}, \mu M^b$			
compd	formula	mp, °C	anal.ª	5-HETE	LTB ₄		
4	$C_{10}H_{12}N_2O_4$	126	C, H, N	18	15		
5	$C_{14}H_{20}N_2O_4$	59	C, H, N	1.2	1.4		
6	$C_{13}H_{16}N_2O_4$	oil	C, H, N	5.0	6.0		
7	$C_{17}H_{18}N_2O_4$	126	C, H, N	2.7	2.9		
16	$C_{16}H_{16}N_2O_5$	146	C, H, N	0.12	0.14		
17	$C_{17}H_{18}N_2O_5$	60	C, H, N	0.098	0.11		
18	$C_{16}H_{15}FN_2O_5$	168	C, H, N	0.10	0.12		
21	$C_{16}H_{16}N_2O_5$	136	$H, N; C^{c}$	0.39	0.45		
22	$C_{17}H_{18}N_2O_5$	150	C, H, N	0.48	0.42		
24	$C_{16}H_{15}FN_2O_5$	173	C, H, N, F	0.085	0.097		
26	$C_{16}H_{15}FN_2O_5$	175	C, H, N, F	0.18	0.17		
zileuton				1.8 ± 0.17	2.2 ± 0.36		

^a Elemental analyses were within $\pm 0.4\%$ of the calculated values unless otherwise noted. The ¹H NMR, ¹³C NMR, IR, and MS data for all the compounds are fully consistent with the assigned structures. ^b In vitro 5-LO inhibitory activity was determined on the basis of A23187-stimulated conversion of [¹⁴C]AA to the 5-LO products in guiena pig peritoneal PMN's. The IC₅₀ values reported are calculated for both 5-HETE and LTB₄ production and represent the means of at least two independent experiments. The SEM values are given for zileuton (n = 3). ^c C: calcd, 60.76; found, 60.34.

were optimal for inhibitory activity. In the present study, potent inhibitors were identified through the same approach. A brief effort was made to investigate inhibitors where the pharmacophore was placed in the center part of the molecule rather than at the termini (compounds 5-7) without much success, though the positive correlation between the potency and overall lipophilicity of the molecule is evident (cf. 4 vs 5-7).²⁰ The position of the aryloxy substituent appears to be critical since the 7-phenoxy isomers (16, 17) are about 3-5 times more potent than the 6-phenoxy isomers (21, 22). We previously observed the same tendency in the benzoxepin series.²¹ The p-fluoro group does not seem to exert any significant effects on intrinsic potency (cf. 16 vs 18). Due to the ex vivo efficacy of the 7-pfluorophenoxy derivative 18 as compared to the phenoxy analog 16 (see below), both enantiomers of 18 were prepared with their optical purity exceeding 95% (see the Chemistry section). The negative enantiomer (24)was found to be twice as active as the its optical isomer (26) in this assay.

Selected compounds were subsequently evaluated following a single intravenous (iv) or oral (po) dose in an *ex vivo* dog model,¹³ which was used to determine *in vivo* 5-LO inhibition using *ex vivo* stimulation of whole blood with the calcium ionophore, A23187. The method allows for the calculation of the time that the test

Table 2. Ex Vivo Inhibition of 5-LO Following Intravenous or Oral Administration in Dog

			inhibition, % ^c								
compd	dose, mg/kg ^a	n^{b}	0.08 h	0.25 h	0.5 h	1.0 h	3.0 h	6.0 h	9.0 h	24 h	DA, h^d
5	1.0 (iv)	1	100	100	64	45	12	0	ND ^e	NDe	0.7
1 6	1.0 (iv)	1	100	100	100	100	92	22	0	4	4.0
17	1.0 (iv)	1	100	100	98	74	19	0	9	0	1.5
18	1.0 (iv)	2	94	98	94	92	91	63	34	17	8.2
	1.0 (po)	2	ND^e	87	94	97	97	90	48	38	8.5
21	1.0 (iv)	1	100	100	100	98	14	1	9	2	1.7
24	1.0 (iv)	1	100	94	92	92	100	76	46	0	8.4
	0.3 (po)	2	ND^e	52	79	76	35	43	0	0	3.5
	1.0 (po)	2	ND^e	80	100	100	100	85	45	7	8.4
	3.0 (po)	2	ND^e	100	100	100	100	97	91	40	19
26	1.0 (iv)	1	100	100	100	100	84	15	0	0	3.8
	1.0 (po)	2	ND^e	64	95	83	80	19	20	3	3.7
zileuton	1.0 (iv)	3	96	94	91	87	81	56	47	27	8.5 ± 0.6
	1.0 (po)	3	ND^e	89	94	91	87	74	49	25	9.8 ± 1.2

^a Test compounds and zileuton were administered either intravenously (iv) or orally (po) at the dose specified. ^b n = number of experiments. ^c Ex vivo inhibition was determined by the measurement of A23187-stimulated LTB₄ production in dog blood at the time specified following administration of the drug. ^d Duration of action (DA) was defined as the period of time from maximum inhibition of A23187-stimulated LTB₄ formation to the time point where 50% of maximum inhibition is reached. The DA was determined graphically from a plot of percent inhibition vs time. The SEM values are given for zileuton. ^e ND = not determined.

compounds are biologically active in the circulation, which is a function of plasma clearance or inactivation. The product, LTB_4 , was quantitated by radioimmunoassay. Inhibition of A23187-stimulated LTB_4 formation was calculated as the change in LTB_4 at each time point following compound administration (iv or po) compared to the value of LTB_4 before dosing. The duration of action (DA) following iv or po administration was defined as the period of time from maximum inhibition of A23187-stimulated LTB_4 formation to the time point where 50% of maximum inhibition is reached. The DA was determined graphically from a plot of percent inhibition of A23187-stimulated LTB₄ formation vs time.

Compound 16, to our disappointment, showed rather modest DA of 4.0 h following iv administration of 1.0 mg/kg in the ex vivo dog model. Its N'-methyl analog (17) was even shorter acting (1.5 h). However, placement of the *p*-fluoro group to the distal aryloxy substituent (compound 18) led to a significant increase in the intravenous DA (8.2 h) at 1.0 mg/kg. The data suggests that, at least in dog, p-hydroxylation of the phenoxy ring could be a significant metabolic pathway in vivo, although further studies are needed to clarify this aspect. Upon oral dosing, 18 exhibited the DA of 8.5 h with sustained maximal inhibition at earlier time points. The 6-phenoxy regioisomer 21 and the nonaryloxy-substituted derivative 5 were less active in the ex vivo study, probably due to weaker enzyme inhibitory potency as demonstrated in the *in vitro* assay (see above).

In accord with the aforementioned in vitro studies, the 2S-isomer 24 was considerably more potent following iv and po administration than its enantiomer (26). Compound 24 was therefore selected for further examination. The *ex vivo* inhibition of 5-LO was found to be dose dependent (Table 2). The inhibitory potency compares favorably with zileuton in our dog model.

In conclusion, we have identified potent, orally active N-hydroxyurea inhibitors of 5-LO based on the 1,4benzodioxan template (18, CGS 25667) employing the *in vitro* guinea pig PMN 5-LO assay and the *ex vivo* dog model. In vitro and *in vivo* inhibitory potency was found to be dependent on the absolute stereochemistry, indicated by the fact that the 2S-enantiomer (24, CGS 25997) is twice as active as its antipode (26, CGS 25998). A general method for the preparation of unsymmetrically substituted benzodioxans from salicyl aldehydes is disclosed. Use of optically active glycidyl derivatives allows chiral synthesis of such 1,4-benzodioxans in high optical purity.

Experimental Section

General. Melting points were taken on a Thomas-Hoover capillary melting point apparatus and are uncorrected. ¹H NMR spectra were recorded on a Brucker AC-250 or Brucker AC-300 spectrometer. A Brucker AC-300 spectrometer was used for ¹³C NMR spectra. IR and MS spectra were measured on a Nicolet 5SXB FTIR spectrometer and a Hewlett-Packard GC/MS 5985B spectrometer, respectively. Microanalyses were carried out at Robertson Laboratory, Inc., Madison, NJ.

DMF, dioxane, methylene chloride, and acetonitrile used were all of anhydrous grade obtained from Aldrich Chemicals. THF was distilled from benzophenone ketyl just prior to use. All the other chemicals were from commercial suppliers and used as received, unless otherwise mentioned. As a general rule, all the reactions were carried out under an inert atmosphere.

Biological assay methods are described previously in full detail.¹³

2-[[N-(tert-Butylcarbonyl)-N-[(tert-butylcarbonyl)oxy]amino]methyl]-1,3-benzodioxan (2). 2-(Hydroxymethyl)-1,4-benzodioxan (1; 17.9 g, 107 mmol) was taken in acetonitrile (200 mL) and treated with dibromotriphenylphosphorane (50 g, 119 mmol). The mixture was stirred for 30 min and evaporated. The residue was taken up in a 1:1 mixture of ether/hexane, and triphenylphosphine oxide was removed by filtration. The solvent was removed by evaporation to give 2-(bromomethyl)-1,4-benzodioxan (24.7 g, 67%), contaminated with a small amount of triphenylphosphine oxide (~5%), as a brownish oil. This material was used without further purification: ¹H NMR (250 MHz, CDCl₃) δ 3.70 (dd, J = 5.2and 11.0 Hz, 1 H), 3.77 (dd, J = 3.2 and 11.0 Hz, 1 H), 4.20 (dd, J = 3.6 and 8.4 Hz, 1 H), 4.36 (dd, J = 1.0 and 8.4 Hz, 1 H), 4.42 (m, 1 H), 6.99 (m, 4 H).

N,*O*-bis-(*tert*-Butoxycarbonyl)hydroxylamine (14.0 g, 61 mmol) was dissolved in DMF (150 mL) and treated with sodium hydride (2.64 g, 60% in mineral oil, 66 mmol). The mixture was stirred for 30 min. To this was added 2-(bromomethyl)-1,4-dioxane (10.6 g, 46 mmol) in DMF (30 mL), and the mixture was stirred overnight at room temperature. This was diluted with ether and washed with saturated aqueous ammonium chloride solution. The organic layer was dried over magnesium sulfate and evaporated. The residue was purified by silica gel flash chromatography (3% ethyl acetate in hexane) to give 2 (10.2 g, 58%) as a pale yellow oil: ¹H NMR (250 MHz, CDCl₃) δ 1.48 (s, 9 H), 1.52 (s, 9H), 3.90 (m, 2 H), 4.09 (dd, *J* = 6.0 and 8.9 Hz, 1 H), 4.85 (dd, *J* = 1.0 and 8.9 Hz, 1 H), 4.42 (m, 1 H), 6.68 (m, 4 H).

2-[(*N*-**Hydroxyamino**)**methyl**]-**1,4-benzodioxan (3).** The protected hydroxylamine **2** (2.0 g, 5.3 mmol) was taken in 20 mL of methylene chloride and treated with trifluoroacetic acid (2.45 g, 21.5 mmol) at room temperature for 30 min. The excess trifluoroacetic acid was evaporated, and the residue was diluted with ether. The organic phase was washed twice with aqueous 1 N sodium hydroxide, dried over magnesium sulfate, and evaporated to give 3 (0.86 g, 91%) as a colorless oil: ¹H NMR (250 MHz, DMSO- d_6) δ 2.92 (dd, J = 5.2 and 11.9 Hz, 1 H), 3.02 (dd, J = 4.0 and 11.9 Hz, 1 H), 3.97 (dd, J = 5.2 and 7.2 Hz, 1 H), 4.28 (m, 1 H), 4.36 (dd, J = 1.0 and 7.2 Hz, 1 H), 6.78 (m, 4 H), 7.47 (br s, 1 H).

2-[[N-(Aminocarbonyl)-N-hydroxyamino]methyl]-1,4benzodioxan (4). The hydroxylamine 3 (950 mg, 5.3 mmol) was taken in 1,4-dioxane (20 mL) and treated with trimethylsilyl isocyanate (910 mg, 7.1 mmol). The mixture was heated at reflux for 30 min and cooled to room temperature. The mixture was diluted with saturated aqueous ammonium chloride and extracted with ethyl acetate. The combined organic layer was dried over magnesium sulfate and evaporated. The residue was crystallized from ether/hexane to give 4 (0.70 g, 60%) as a colorless solid: mp 126 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 3.52 (dd, J = 6.5 and 14.2 Hz, 1 H), 3.69 (dd, J = 5.6 and 14.2 Hz, 1 H), 3.98 (dd, J = 6.9 and 11.9 Hz.1 H), 4.31 (dd, J = 2.2 and 11.9 Hz, 1 H), 4.35 (m, 1 H), 6.43(br s, 2 H), 6.82 (m, 4 H), 9.57 (s, 1 H); ¹³C NMR (75.47 MHz, DMSO- d_6 δ 161.47, 142.99, 142.82, 121.39, 121.15, 117.14, 116.89, 70.54, 65.81, 50.15; IR (KBr) 1653, 1594, 1578, 1037, 750 cm⁻¹; MS (CI/CH₄) 225 (M + 1). Anal. ($C_{10}H_{12}N_2O_4$) C, H.N.

2-[[N-[(N'-Butylamino)carbonyl]-N-hydroxyamino]methyl]-1,4-benzodioxan (5). 2-[(N-Hydroxyamino)methyl]-1,4-benzodioxan (3; 860 mg, 4.75 mmol) was dissolved in 20 mL of 1,4-dioxane and treated with butyl isocyanate (710 mg, 7.1 mmol). The mixture was heated at reflux for 30 min and cooled. The mixture was diluted with ether and washed with saturated aqueous ammonium chloride solution. The organic layer was dried over magnesium sulfate and evaporated to give a mixture (1.24 g) of 5 and 2-[[N-[(N'-butylamino)carbonyl]-N-[[(N"-butylamino)carbonyl]oxy]amino]methyl]-1,4-dioxane as an oil. This was not fully characterized and used as obtained in the next step. The crude material was dissolved in a 1:1 mixture of water/2-propanol and treated with lithium hydroxide monohydrate (1.38 g, 32.8 mmol) for 30 min. The mixture was made acidic with 1 N hydrochloric acid and diluted with ethyl acetate. The organic layer was washed with saturated aqueous ammonium chloride solution, dried over magnesium sulfate, and evaporated. Silica gel flash chromatography (2:1 hexane/ethyl acetate) gave 5 (0.35 g, 26%) as a colorless solid: mp 59 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 0.86 (t, J = 7.2 Hz, 3 H), 1.26 (sextet, J = 7.2 Hz, 2 H), 1.40(quintet, J = 7.2 Hz, 2 H), 3.03 (dt, J = 5.8 and 7.2 Hz, 2 H), 3.49 (dd, J = 6.4 and 14.1 Hz, 1 H), 3.64 (dd, J = 5.9 and 14.1Hz, 1 H), 3.98 (dd, J = 7.0 and 11.6 Hz, 1 H), 4.30 (dd, J = 2.1and 11.6 Hz, 1 H), 4.35 (m, 1 H), 6.82 (m, 4 H), 7.01 (t, J = 5.8 Hz, 1 H), 9.50 (s, 1 H); ¹³C NMR (75.47 MHz, DMSO- d_6) δ 160.66, 142.99, 142.82, 121.39, 121.14, 117.12, 116.89, 70.53, 65.83, 50.92, 38.99, 31.90, 19.47, 13.72; IR (KBr) 1634, 1592, 1547, 1263, 740 cm⁻¹; MS (CI/CH₄) 281 (M + 1). Anal. $(C_{14}H_{20}N_2O_4)$ C, H, N.

2-[[N-[(N '-Benzylamino)carbonyl]-N-hydroxyamino]-methyl]-1,4-benzodioxan (6). This compound was prepared from **3** by the reaction with allyl isocyanate according to the procedure described for **5** and isolated as a tan solid: mp 120 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 3.52 (dd, J = 6.4 and 14.3 Hz, 1 H), 3.6-3.8 (m, 3 H), 3.98 (dd, J = 7.0 and 11.5 Hz, 1 H), 4.31 (dd, J = 2.2 and 11.5 Hz, 1 H), 4.37 (m, 1 H), 5.05 (dd, J = 1.5 and 10.3 Hz, 1 H), 5.10 (dd, J = 1.5 and 17.2 Hz, 1 H), 5.79 (m, 1 H), 6.83 (m, 4 H), 7.17 (t, J = 5.9 Hz, 1 H), 9.58 (s, 1 H); ¹³C NMR (75.47 MHz, DMSO- d_6) δ 160.45, 142.98, 142.81, 136.30, 121.39, 121.14, 117.14, 116.88, 114.54, 70.47, 65.80, 54.88, 50.84; IR (KBr) 1657, 1649, 1594, 1530, 1275, 1227 cm⁻¹; MS (CL/CH₄) 264 (M + 1). Anal. (C₁₃H₁₆N₂O₄) C, H, N.

2-[[N-[(N'-Allylamino)carbonyl]-N-hydroxyamino]m-ethyl]-1,4-benzodioxan (7). This compound was prepared from 3 by the reaction with benzyl isocyanate according to the

procedure described for **5** and isolated as a pale yellow oil: ¹H NMR (300 MHz, DMSO- d_6) δ 3.53 (dd, J = 6.3 and 14.3 Hz, 1 H), 3.74 (dd, J = 5.0 and 14.3 Hz, 1 H), 3.99 (dd, J = 7.0 and 11.5 Hz, 1 H), 4.24 (d, J = 6.3 Hz, 2 H), 4.30 (dd, J = 2.2 and 11.5 Hz, 1 H), 4.37 (m, 1 H), 6.81 (m, 4 H), 7.1–7.3 (m, 5 H), 7.61 (t, J = 6.3 Hz, 1 H), 9.61 (s, 1 H); ¹³C NMR (75.47 MHz, DMSO- d_6) δ 160.66, 142.98, 142.79, 140.52, 128.10, 127.14, 126.55, 121.39, 121.14, 117.15, 116.89, 70.54, 65.78, 50.83, 42.94; IR (KBr) 1635, 1539, 1264 cm⁻¹; MS (CI/CH₄) 315 (M + 1). Anal. (C₁₇H₁₈N₂O₄) C, H, N.

2-[(2,3-Oxopropyl)oxy]-5-phenoxybenzaldehyde (10). 2-Hydroxy-5-phenoxybenzaldehyde (8;13 4.23 g, 19.8 mmol) and cesium carbonate (32.2 g, 98.8 mmol) were taken in DMF (200 mL) and treated with epibromohydrin (3.25 g, 23.7 mmol). The mixture was stirred overnight at room temperature. This was diluted with ether, washed with water and brine, dried over magnesium sulfate, and evaporated. Purification by silica gel flash chromatography (4:1 hexane/ethyl acetate) gave 10 (4.18 g, 78%) as a colorless oil: ¹H NMR (300 MHz, $CDCl_3$) δ 2.79 (dd, J = 2.6 and 4.5 Hz, 1 H), 2.95 (t, J = 4.5 Hz, 1 H), 3.41 (m, 1 H), 4.04 (dd, J = 5.9 and 11.2 Hz, 1 H), 4.40 (dd, J = 2.7and 11.2 Hz, 1 H), 6.95 (d, J = 8.1 Hz, 2 H), 6.99 (d, J = 9.0Hz, 1 H), 7.09 (t, J = 7.4 Hz, 1 H), 7.24 (dd, J = 3.1 and 9.0 Hz, 1 H), 7.31 (t, J = 8.0 Hz, 2 H), 7.47 (d, J = 3.1 Hz, 1 H); IR (CH₂Cl₂) 2717, 1685, 1591, 1488, 1215, 836, 817 cm⁻¹; MS (CI/CH_4) 271 (M + 1).

2-(Hydroxymethyl)-7-phenoxy-1,4-benzodioxan (12). The epoxy aldehyde 10 (4.5 g, 11.6 mmol) was dissolved in chloroform (100 mL) and treated with m-chloroperbenzoic acid (4.36 g, 80%, 25.2 mmol). The mixture was heated at reflux for 3 h. Ether was added, and the organic phase was washed three times with saturated aqueous sodium bicarbonate solution, dried over magnesium sulfate, and evaporated. Purification with silica gel flash chromatography (10% ethyl acetate/ hexane) gave 2-[(2,3-oxopropyl)oxy]-5-phenoxyphenol (2.76 g, 69%) as a colorless oil: ¹H NMR (300 MHz, CDCl₃) δ 2.75 (dd, J = 2.5 and 4.5 Hz, 1 H), 2.87 (t, J = 4.5 Hz, 1 H), 3.25 (m, 1 H), 3.96 (dd, J = 6.0 and 11.2 Hz, 1 H), 4.19 (dd, J = 2.5 and11.2 Hz, 1 H), 6.26 (s, 1 H), 6.40 (dd, J = 2.7 and 8.8 Hz, 1 H), 6.58 (d, J = 2.7 Hz, 1 H), 6.77 (d, J = 8.8 Hz, 1 H), 6.90 (d, J)= 8.0 Hz, 2 H), 6.99 (t, J = 8.0 Hz, 1 H), 7.22 (t, J = 8.0 Hz)2 H

The phenol thus formed (2.76 g, 10.7 mmol) was taken in 1,4-dioxane (30 mL) and treated with aqueous 1 N sodium hydroxide (22 mmol). The reaction mixture was stirred at room temperature for 2 h. The mixture was diluted with ether, extracted with saturated aqueous sodium chloride solution, dried over magnesium sulfate, and evaporated. Purification with silica gel flash chromatography (1:2 ethyl acetate/hexane) gave 12 (2.76 g, 73%) as a colorless oil: ¹H NMR (300 MHz, CDCl₃) δ 3.83 (dd, J = 5.2 and 11.2 Hz, 1 H), 3.90 (dd, J = 4.2 and 11.2 Hz, 1 H), 4.11 (m, 1 H), 4.29 (m, 2 H), 6.55 (dd, J = 2.7 and 8.8 Hz, 1 H), 6.60 (d, J = 2.7 Hz, 1 H), 6.85 (d, J = 8.8 Hz, 1 H), 6.97 (d, J = 7.7 Hz, 2 H), 7.06 (t, J = 7.7 Hz, 1 H), 7.31 (t, J = 7.7 Hz, 2 H); IR (film) 1594, 1217, 1146, 1139 cm⁻¹.

2-[(N-Hydroxyamino)methyl]-7-phenoxy-1,4-benzodioxan (14). The cyclization product 12 (1.42 g, 5.50 mmol) and triethylamine (1.11 g, 11.0 mmol) were dissolved in methylene chloride (50 mL) and treated with methanesulfonyl chloride (0.94 g, 8.26 mmol) at 0 °C. The mixture was stirred at 0 °C for 30 min and diluted with ether. This was washed with aqueous sodium bicarbonate, aqueous 1 N hydrochloric acid, and saturated aqueous ammonium chloride, dried over magnesium sulfate, and evaporated. The residue was purified by silica gel flash chromatography (1:3 ethyl acetate/hexane) to give 2-[[(methylsulfonyl)oxy]methyl]-7-phenoxy-1,4-benzodioxan (1.85 g, 100%) as a tan oil which partially solidified upon standing: ¹H NMR (300 MHz, CDCl₃) & 3.06 (s, 3 H), 4.10 (dd, J = 6.0 and 11.7 Hz, 1 H), 4.29 (dd, J = 4.2 and 11.7 Hz, 1 H), $4.4-4.6 \text{ (m, 3 H)}, 6.55 \text{ (m, 2 H)}, 6.84 \text{ (d, } J = 8.7 \text{ Hz}, 1 \text{ H)}, 6.95 \text{ (m, 2 H)}, 6.95 \text{ (m, 3 H)}, 6.95 \text$ (d, J = 7.7 Hz, 2 H), 7.05 (t, J = 7.7 Hz, 1 H), 7.30 (t, J = 7.7 Hz)Hz, 2 H); IR (film) 1594, 1501, 1365, 1217, 1147 cm⁻¹; MS (CI/ CH_4) 337 (M + 1).

The mesylate (1.85 g, 5.5 mmol) and sodium iodide (1.54 g, 11.0 mmol) were taken in methyl ethyl ketone (60 mL) and heated at reflux for 3 h. The mixture was cooled and diluted with ethyl acetate. This was washed with aqueous sodium

bicarbonate, dried over magnesium sulfate, and evaporated. Purification of the residue by silica gel flash chromatography (hexane) gave 14 (1.83 g, 90%) as a tan oil: ¹H NMR (300 MHz, CDCl₃) δ 3.32 (m, 2 H), 4.12 (dd, J = 5.8 and 11.1 Hz, 1 H), 4.26 (m, 1 H), 4.34 (dd, J = 2.2 and 11.1 Hz, 1 H), 6.53 (dd, J =2.7 and 8.7 Hz, 1 H), 6.59 (d, J = 2.7 Hz, 1 H), 6.83 (d, J =8.7 Hz, 1 H), 6.96 (d, J = 7.7 Hz, 2 H), 7.05 (d, J = 7.7 Hz, 1 H), 7.29 (t, J = 7.7 Hz, 2 H); IR (film) 1595, 1500, 1285 cm⁻¹; MS (CI/CH₄) 369 (M + 1).

2-[[(N-(Aminocarbonyl)-N-hydroxyamino]methyl]-7phenoxy-1,4-benzodioxan (16). This compound was obtained as a colorless solid from **14** according to the procedure described for 4: mp 146 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 3.51 (dd, J = 6.4 and 14.4 Hz, 1 H), 3.69 (dd, J = 5.9 and 14.4 Hz, 1 H), 3.98 (dd, J = 5.8 and 11.6 Hz, 1 H), 4.31 (dd, J = 2.1and 11.6 Hz, 1 H), 4.35 (m, 1 H), 6.42 (br s, 2 H), 6.52 (m, 2 H), 6.89 (d, J = 9.5 Hz, 1 H), 6.94 (d, J = 7.4 Hz, 2 H), 7.08 (t, J = 7.4 Hz, 1 H), 7.35 (t, J = 7.4 Hz, 2 H), 9.55 (s, 1 H); ¹³C NMR (75.47 MHz, DMSO- d_6) δ 161.44, 157.48, 150.21, 143.33, 139.30, 129.90, 122.91, 117.80, 117.56, 112.02, 108.20, 70.81, 65.66, 50.05; IR (KBr) 1648, 1217 cm⁻¹; MS (CI/CH₄) 317 (M + 1). Anal. (C₁₆H₁₆N₂O₅) C, H, N.

 $\label{eq:linear} 2-[[N-[(N'-Methylamino)carbonyl]-N-hydroxyamino]$ methyl]-7-phenoxy-1,4-benzodioxan (17). 2-[(N-Hydroxyamino)methyl]-7-phenoxy-1,4-benzodioxan (14; 200 mg, 0.73 mmol) was taken in acetonitrile (10 mL) and treated with methyl isocyanate (42 mg, 0.73 mmol) at room temperature. This was stirred for 3 h at room temperature and diluted with ethyl acetate. The organic phase was washed with saturated aqueous ammonium chloride, dried over magnesium sulfate, and evaporated. Crystallization of the residue from ethyl acetate/hexane gave the title compound 17 (93 mg, 38%) as a glassy solid: mp 60 °C; ¹H NMR (300 MHz, DMSO-d₆) & 2.59 $(d, J = 3.6 \text{ Hz}, \overline{3} \text{ H}), 3.49 (dd, J = 6.2 \text{ and } 14.3 \text{ Hz}, 1 \text{ H}), 3.70$ (dd, J = 6.2 and 14.3 Hz, 1 H), 3.99 (dd, J = 6.9 and 11.6 Hz,1 H), 4.30 (dd, J = 2.2 and 11.6 Hz, 1 H), 4.37 (d of quartet, J= 2.2 and ~ 6.5 Hz, 1 H), 6.5 (m, 2 H), 6.8-7.0 (m, 4 H), 7.08 $(t, J = 7.4 \text{ Hz}, 1 \text{ H}), 7.34 (t, J = 7.4 \text{ Hz}, 2 \text{ H}), 9.49 (s, 1 \text{ H}); {}^{13}\text{C}$ NMR (75.47 MHz, DMSO- d_6) δ 161.21, 157.49, 150.18, 143.30, 139.30, 129.89, 122.89, 117.77, 117.56, 112.03, 108.24, 70.77, 65.62, 50.84, 26.51; IR (KBr) 1645, 1541, 1500, 1216, 1251, 1145 cm⁻¹; MS (CI/CH₄) 331 (M + 1). Anal. ($C_{17}H_{18}N_2O_5$) C, H, N.

2-[[N-(Aminocarbonyl)-N-hydroxyamino]methyl]-7-(4-fluorophenoxy)-1,4-benzodioxan (18). This compound was obtained as a colorless solid from **15** according to the procedure described for 4: mp 168 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 3.51 (dd, J = 6.4 and 14.3 Hz, 1 H), 3.69 (dd, J = 5.9 and 14.3 Hz, 1 H), 3.89 (dd, J = 6.9 and 11.6 Hz, 1 H), 4.30 (dd, J = 2.0 and 11.6 Hz, 1 H), 4.35 (m, 1 H), 6.43 (br s, 2 H), 6.50 (m, 2 H), 6.88 (d, J = 9.4 Hz, 1 H), 9.55 (s, 1 H); ¹³C NMR (75.47 MHz, DMSO- d_6) δ 161.36, 157.77 ($^{J}J_{CF} = 238.8$ Hz), 153.37 ($^{4}J_{C.F} = 2.2$ Hz), 150.59, 143.23, 139.13, 119.62 ($^{3}J_{C.F} = 8.4$ Hz), 117.49, 116.30 ($^{2}J_{C.F} = 23.4$ Hz), 111.47, 70.70, 65.54, 49.93; IR (KBr) 1662, 1615, 1595, 1575, 1251, 1191, 1142 cm⁻¹; MS (CI/CH₄) 335 (M + 1). Anal. (C₁₆H₁₆FN₂O₆) C, H, N.

2-[(**2**,**3-Oxopropy**])**oxy**]-**4-phenoxypheny**] **Benzy**] **Ether** (**19**). 5-Phenoxy-2-hydroxybenzaldehyde (**8**; 20 g, 94 mmol) and potassium carbonate (26.1 g, 188 mmol) were taken in DMF (250 mL) and treated with benzyl bromide (16.1 g, 94 mmol). The mixture was stirred at room temperature overnight. This was diluted with ethyl acetate, washed three times with saturated aqueous ammonium chloride, dried over magnesium sulfate, and evaporated to give 2-(benzyloxy)-5-phenoxybenzaldehyde (25.2 g, 88%) as a colorless solid. This was used without further purification: mp 64-65 °C; ¹H NMR (250 MHz, CDCl₃) δ 5.17 (s, 2 H), 6.93 (d, J = 7.6 Hz, 2 H), 7.03 (d, J = 8.9 Hz, 2 H), 7.09 (d, J = 7.6 Hz, 1 H), 7.2-7.6 (m, 8 H), 10.48 (s, 1 H).

The benzyl ether (22.25 g, 73 mmol) was dissolved in methanol (200 mL) and treated with hydrogen peroxide (31%in water, 32.0 mL) and concentrated sulfuric acid (15.0 mL). The mixture was stirred overnight at room temperature and evaporated. The residue was dissolved in saturated aqueous sodium chloride solution and extracted with ether. The organic phase was dried over magnesium sulfate and evaporated. Purification of the residue by silica gel flash chromatography (5% ethyl acetate/hexane) gave 2-(benzyloxy)-5-phenoxyphenol (16.5 g, 77%) as a colorless solid: mp 36-37 °C; ¹H NMR (250 MHz, CDCl₃) δ 5.07 (s, 2 H), 5.71 (s, 1 H), 6.44 (dd, J = 2.8 and 8.8 Hz, 1 H), 6.61 (d, J = 2.8 Hz, 1 H), 6.84 (d, J = 8.8 Hz, 1 H), 6.9-7.1 (m, 5 H), 7.39 (m, 5 H).

The phenol (16 g, 55 mmol) and cesium carbonate (26.9 g, 82.5 mmol) were taken in DMF (200 mL) and treated with epibromohydrin (11.2 g, 83 mmol). The mixture was stirred at room temperature for 2 h and diluted with ether. The organic phase was washed several times with saturated aqueous sodium chloride solution, dried over magnesium sulfate, and evaporated. Purification of the residue by silica gel flash chromatography (10% ethyl acetate/hexane) gave **19** (14.9 g, 78%) as a colorless oil: ¹H NMR (250 MHz, CDCl₃) δ 2.72 (dd, J = 2.6 and 4.7 Hz, 1 H), 2.86 (t, J = 4.7 Hz, 1 H), 3.94 (m, 1 H), 3.96 (dd, J = 5.6 and 11.3 Hz, 1 H), 4.22 (dd, J = 2.8 and 11.3 Hz, 1 H), 5.08 (s, 2 H), 6.47 (dd, J = 2.8 and 8.7 Hz, 1 H), 6.64 (d, J = 2.8 Hz, 1 H), 6.85 (d, J = 8.7 Hz, 1 H), 6.9–7.1 (m, 5 H), 7.3–7.5 (m, 5 H).

2-(Hydroxymethyl)-6-phenoxy-1,4-benzodioxan (20). The benzyl ether **19** (3.68 g, 10.6 mmol) and palladium on charcoal (10%, 1.84 g) were taken in ethyl acetate and subjected to hydrogenation at atmospheric pressure. The reaction was terminated in 15 min. The catalyst was removed by filtration over a bed of Celite, and the solvent was evaporated. Purification of the residue by silica gel flash chromatography (15% ethyl acetate/hexane) gave 2-[(2,3-oxopropyl)oxy]-4-phenoxyphenol (2.19 g, 50%) as a colorless oil: ¹H NMR (250 MHz, CDCl₃) δ 2.76 (dd, J = 2.6 and 4.6 Hz, 1 H), 2.90 (t, J = 4.6 Hz, 1 H), 3.34 (m, 1 H), 3.93 (dd, J = 5.7 and 11.3 Hz, 1 H), 4.26 (dd, J = 2.7 and 11.3 Hz, 1 H), 6.59 (d, J = 2.6 Hz, 1 H),

The resultant phenol (6.0 g, 23 mmol) was taken in ethanol (250 mL) and treated with aqueous 1 N sodium hydroxide (23 mL). The mixture was stirred for 30 min and diluted with ether. The organic phase was washed with 1 N hydrochloric acid and water, dried over magnesium sulfate, and evaporated. Purification of the residue by silica gel flash chromatography (25% ethyl acetate/hexane) gave **20** (4.29 g, 71%) as a colorless oil: ¹H NMR (250 MHz, CDCl₃) δ 3.82 (dd, J = 4.7 and 12.0 Hz, 1 H), 3.91 (dd, J = 3.8 and 12.0 Hz, 1 H), 4.09 (dd, J = 7.5 and 10.7 Hz, 1 H), 4.22 (m, 1 H), 4.28 (dd, J = 1.6 and 10.7 Hz, 1 H), 5.62 (m, 2 H), 6.83 (d, J = 8.5 Hz, 1 H), 6.85–7.05 (m, 5 H).

2-[(N-Hydroxyamino)methyl]-6-phenoxy-1,4-benzodioxan (21). This compound was prepared from **20** according to the procedure described for **16** and obtained as a colorless solid: mp 136 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 3.53 (dd, J= 6.4 and 14.3 Hz, 1 H), 3.70 (dd, J = 5.6 and 14.3 Hz, 1 H), 4.00 (dd, J = 7.4 and 11.8 Hz, 1 H), 4.32 (m, 2 H), 6.44 (br s, 2 H), 6.51 (dd, J = 2.8 and 8.7 Hz, 1 H), 6.56 (d, J = 2.8 Hz, 1 H), 6.87 (d, J = 8.7 Hz, 1 H), 6.94 (d, J = 7.6 Hz, 2 H), 7.07 (t, J = 7.6 Hz, 1 H), 7.34 (t, J = 7.6 Hz, 2 H), 9.57 (s, 1 H); ¹³C NMR (75.47 MHz, DMSO- d_6) δ 161.47, 157.50, 149.94, 143.46, 139.15, 129.89, 122.87, 117.79, 117.72, 112.29, 108.05, 70.43, 66.00, 50.10; IR (KBr) 1667, 1590, 1501, 1232, 1216, 1143 cm⁻¹; MS (CI/CH₄) 317 (M + 1). Anal. (C₁₆H₁₆N₂O₅) C, H, N.

2-[[N-[(N'-Methylamino)carbonyl]-N-hydroxyamino]methyl]-6-phenoxy-1,4-benzodioxan (22). This compound was prepared according to the procedure described for **17** and obtained as a colorless solid: mp 150 °C; ¹H NMR (300 MHz, DMSO- d_6) δ 2.60 (d, J = 4.6 Hz, 3 H), 3.50 (dd, J = 6.4 and 14.2 Hz, 1 H), 3.70 (dd, J = 5.7 and 14.2 Hz, 1 H), 4.00 (dd, J= 7.4 and 11.8 Hz, 1 H), 4.3 (m, 2 H), 6.51 (dd, J = 2.8 and 8.7 Hz, 1 H), 6.56 (d, J = 2.8 Hz, 1 H), 6.86 (d, J = 8.7 Hz, 1 H), 6.93 (d, J = 7.5 Hz, 2 H), 7.00 (quartet, J = 4.6 Hz, 1 H), 7.07 (t, J = 7.5 Hz, 2 H), 7.00 (quartet, J = 4.6 Hz, 1 H), 7.07 (t, J = 7.5 Hz, 2 H), 7.08 (t, J = 7.5 Hz, 2 H), 9.50 (s, 1 H); ¹³C NMR (75.47 MHz, DMSO- d_6) δ 161.22, 157.51, 149.94, 143.46, 139.15, 129.89, 122.87, 117.80, 117.71, 112.31, 108.06, 70.43, 65.99, 50.93, 26.53; IR (KBr) 1653, 1649, 1211, 1147 cm⁻¹; MS (CI/CH₄) 331 (M + 1). Anal. (C₁₇H₁₈N₂O₅) H, N; C: calcd, 60.76; found, 60.34.

(2S)-(-)-2-[[N-(Aminocarbonyl)-N-hydroxyamino]methyl]-7-(4-fluorophenoxy)-1,4-benzodioxan (24). This compound was prepared according to the procedure described for 16, using (2R)-(-)-glycidyl tosylate (23) and 9 as the starting materials: mp 173 °C; $[\alpha]_D$ -46.50° (c 0.92, DMSO). Anal. $(C_{16}H_{15}FN_2O_5)$ C, H, N, F.

(2R)-(+)-2-[[N-(Aminocarbonyl)-N-hydroxyamino]methyl]-7-(4-fluorophenoxy)-1,4-benzodioxan (26). This compound was prepared according to the procedure described for 16, using (2S)-(+)-glycidyl tosylate (25) and 9 as the starting materials: mp 175 °C; $[\alpha]_D$ +39.09° (c 0.74, DMSO). Anal. (C₁₆H₁₅FN₂O₅) C, H, N, F.

Acknowledgment. We appreciate continuous encouragement and helpful discussion by Drs. Jim Stanton and Dennis Roland. We also thank our Analytical Staff, particularly Mr. Karl Gunderson, Mr. Mike Hatolski, Ms. Natalie Cahoon, Ms. Lia Raabis, Mr. Stu Brody, and Ms. Magda Brzechffa, for the analytical data and the members of Scientific Information Center, including Ms. Ellen Donoghue, Ms. Debbie Juterbock, Ms. Carole Coppola, and Ms. Barbara Ikler, for excellent literature search service.

References

- Leukotrienes and Lipoxygenases; Rokach, J., Ed.; Elsevier: New York, 1989.
- For leading references, see: (a) Batt, D. G. 5-Lipoxygenase Inhibitors and Their Anti-Inflammatory Activities. Prog. Med. Chem. 1992, 29, 1-63. (b) Musser, J. H.; Kreft, A. F. 5-Lipoxygenase: Properties, Pharmacology, and The Quinolinyl(bridged)aryl Class of Inhibitors. J. Med. Chem. 1992, 35, 2501-2524.
 (a) Carter, G. W.; Young, P. R.; Albert, D. H.; Bouska, J.; Dyer, R.; Bell, R. L.; Summers, J. B.; Brooks, D. W. 5-Lipoxygenase Content of the State o
- (3) (a) Carter, G. W.; Young, P. R.; Albert, D. H.; Bouska, J.; Dyer, R.; Bell, R. L.; Summers, J. B.; Brooks, D. W. 5-Lipoxygenase Inhibitory Activity of Zileuton. J. Pharmacol. Exp. Ther. 1991, 256, 929-937. (b) Bell, R. L.; Young, P. R.; Albert, D.; Lanni, C.; Summers, J. B.; Brooks, D. W.; Rubin, P.; Carter, G. W. The Discovery and Development of Zileuton: An Orally Active 5-Lipoxygenase Inhibitor. Int. J. Immunopharmacol. 1992, 14, 505-510.
- (4) (a) Gillard, J.; Ford-Hutchinson, A. W.; Chan, C.; Charleson, S.; Denis, D.; Foster, A.; Fortin, R.; Leger, S.; McFarlane, C. S.; Morton, H.; Piechuta, H.; Riendeau, D.; Rouzer, C. A.; Rokach, J.; Young, R.; MacIntyre, D. E.; Peterson, L.; Bach, T.; Eiermann, G.; Hopple, S.; Humes, J.; Hupe, L.; Luell, S.; Metzger, J.; Meurer, R.; Miller, D. K.; Opas, E.; Pacholok, S. L-663,536 (3-(1-(4-Chlorobenzyl)-3-t-butyl-thio-5-isopropylindol-2-yl)-2,2-dimethylpropanoic acid), a Novel, Orally Active Leukotriene Biosynthesis Inhibitor. Can. J. Physiol. Pharmacol. 1989, 67, 456-464. (b) Gilland, J. W.; Dixon, R.; Ethier, D.; Evans, J.; Ford-Hutcinson, A. W.; Fortin, R.; Girard, Y.; Guidon, Y.; Hamel, P.; Jones, T.; Leveillé, C.; Lord, A.; Miller D.; Morton, H.; Rouzer, C.; Yoakim, C. Therapeutic Potential of 5-Lipoxygenase Inhibitors: The Discovery and Development of MK-886, a Novel-Mechanism Leukotriene Inhibitor. Inflammation Drug Ther. Ser. 1992, 5, 275-286. (c) Depre, M.; Friedman, B.; Tanaka, W.; Van Hecken, A.; Buntinx, A.; DeSchepper, P. J. Biochemical Activity, Pharmacokinetics, and Tolerability of MK-886, a Leukotriene Biosynthesis Inhibitor, in Humans. Clin. Pharmacol. Ther. 1993, 53, 602-607.
- (5) (a) Prasit, P.; Belley, M.; Brideau, C.; Chan, C.; Charleson, S.; Evans, J. F.; Fortin, R.; Ford-Hutchinson, A. W.; Gillard, J. W.; Guay, J.; Hutchinson, J. H.; Léger, S.; Riendeau, D.; Young, R. N.; Zamboni, R. A New Class of Leukotriene Biosynthesis Inhibitors: The Discovery of MK0591. Bioorg. Med. Chem. Lett. 1992, 2, 1395-1398. (b) Prasit, P.; Belley, M.; Blouin, M.; Brideau, C.; Chan, C.; Charleson, S.; Evans, J. F.; Frenette, R.; Gauthier, J. Y.; Guay, J.; Gillard, J. W.; Grimm, E.; Ford-Hutchinson, A. W.; Hutchinson, J. H.; Fortin, R.; Jones, T. R.; Mancini, J.; Léger, S.; McFarlane, C. S.; Piechuta, H.; Riendeau, D.; Roy, P.; Tagari, P.; Vickers, P. J.; Young, R. N.; Zamboni, R. A New Class of Leukotriene Biosynthesis Inhibitors: The Discovery of MK-0591. J. Lipid Mediators 1993, 6, 239-244.
 (6) (a) Musser, J. H.; Kreft, A. F. Substituted-(2-quinolynyl(bridged)-
- (6) (a) Musser, J. H.; Kreft, A. F. Substituted-(2-quinolynyl(bridged)aryl)compounds: Modulators of Eicosanoid Biosynthesis and Action. Drugs Future 1990, 15, 73-80. (b) Grimes, D.; Sturm. R. J.; Marinari, L. R.; Carlson, R. P.; Berkendopf, J. W.; Musser, J. H.; Kreft, A. F.; Weichman, B. M. WY-50,295 Tromethamine, a Novel, Orally Active 5-Lipoxygenase Inhibitor: Biochemical Characterization and Antiallergic Activity. Eur. J. Pharmacol. 1993, 236, 217-228. See also ref 2b.
- (7) (a) Crawley, G. C.; Dowell, R. I.; Edwards, P. N.; Foster, S. J.; McMillan, R. M.; Walker, E. R. H.; Waterson, D. Methoxytetrahydropyrans. A New Series of Selective and Orally Potent 5-Lipoxygenase Inhibitors. J. Med. Chem. 1992, 35, 2600-2609.
 (b) McMillan, R. M.; Spruce, K. E.; Crawley, G. C.; Walker, E. R. H.; Foster, S. J. Pre-Clinical Pharmacology of ICI D2138, a Potent Orally-Active Non-Redox Inhibitor of 5-Lipoxygenase. Br. J. Pharmacol. 1992, 107, 1042-1047.

- (8) (a) Weinblatt, M.; Kremer, J.; Helfgott, S.; Coblyn, J.; Maier, A.; Sperling, R.; Petrillo, G.; Kesterson, J.; Dube, L.; Henson, B.; Teoh, N.; Rubin, P. A 5-Lipoxygenase Inhibitor in Rheumatoid Arthritis (RA). Arthritis Rheum. 1990, 33, S152. (b) Israel, E.; Dermarkarian, R.; Rosenberg, M.; Sperling, R.; Taylor, G.; Rubin, P.; Drazen, J. M. The Effects of a 5-Lipoxygenase Inhibitor on Asthma Induced by Cold, Dry Air. N. Engl. J. Med. 1990, 323, 1740-1744. (c) Knapp, H. R. Reduced Allergen-Induced Nasal Congestion and Leukotriene Synthesis with an Orally Active 5-Lipoxygenase Inhibitor. N. Engl. J. Med. 1990, 323, 1745-1748. (d) Staerk Laursen, L.; Naesdal, J.; Bukhave, K.; Lauritsen, K.; Rask-Madsen, J. Selective 5-Lipoxygenase Inhibition in Ulcerative Colitis. Lancet 1990, 335, 683-685. (e) Hui, K. P.; Taylor, I. K.; Taylor, G. W.; Rubin, P.; Kesterson, J.; Barnes, N. C.; Barnes, P. J. Effect of 5-Lipoxygenase Inhibitor on Leukotriene Generation and Airway Responses after Allergen Challenge in Asthmatic Patients. Thorax 1991, 46, 184-189. (f) Busse, W. W.; Gaddy, J. N. The Role of Leukotriene Antagonists and Inhibitors in The Treatment Of Airway Disease. Am. Rev. Respir. Dis. 1991, 143, S103-107. (g) Stenson, W. F.; Lauritsen, K.; Laursen, L. S.; Rask-Madsen, J.; Jacobsen, O.; Naesdal, J.; Cort, D.; Goebell, H.; Pesker, B.; Hanauer, S.; Swanson, L.; Dube, L.; Rubin, P. A Clinical Trial of Zileuton, a Specific Inhibitor of 5-Lipoxygenase, in Ulcerative Colitis. Gastroenterology 1991, 100, A253. (h) Collawn, C.; Rubin, P.; Perez, N.; Bobadilla, J.; Cabrera, G.; Reyes, E.; Borovoy, J.; Kershenobich, D. Phase II Study of the Safety and Efficacy of a 5-Lipoxygenase Inhibitor in Patients with Ulcerative Colitis. Am. J. Gastroenterol. 1992, 87, 342-346. (i) Rask-Madsen, J.; Bukhave, K.; Laursen, L. S.; Lauritsen, K. 5-Lipoxygenase Inhibitors for the Treatment of Inflammatory Bowel Disease. Agents Actions 1992, 35, C37-46. (j) Weinblatt, M. E.; Kermer, J. M.; Coblyn, J. S.; Helfgott, S.; Maier, A. L.; Petrillo, G.; Henson, B.; Rubin, P.; Sperling, R. Zileuton, a 5-Lipoxygenase Inhibitor in Rheumatoid Arthritis. J. Rheumatol. 1992, 19, 1537-1541. (k) Israel, E.; Rubin, P.; Kemp, J. P.; Grossman, J.; Pierson, W.; Siegel, S. C.; Tikelman, D.; Murray, J. J.; Busse, W.; Segal, A. T.; Fish, J.; Kaiser, H. B.; Ledford, D.; Wenzel, S.; Rosenthal, R.; Cohn, J.; Lanni, C.; Pearlman, H.; Karahalios, P.; Drazen, J. M. The Effect of Inhibition of 5-Lipoxygenase by Zileuton in Mild-to-Moderate Asthma. Ann. Intern. Med. 1993, 119, 1059-1066. (l) Israel, E.; Fischer, A. R.; Rosenberg, M. A.; Lilly, C. M.; Callery, J. C.; Shapiro, J.; Cohn, J.; Rubin, P.; Drazen, J. M. The Pivotal Role of 5-Lipoxygenase Products in the Reaction of Aspirin-Sensitive Asthmatics to Aspirin. Am. Rev. Respir. Dis. **1993**, *148*, 1447-1451. (m) Laursen, L. S.; Lauritsen, K.; Bukhave, K.; Rask-Madsen, J.; Jacobsen, O.; Naesdal, J.; Goebell, H.; Peskar, B.; Rubin, P. D.; Swanson, L. J.; Kesterson, J. W.; Keshavarzian, A.; Dozarek, R.; Hanauer, S.; Cort, D.; Stenson, W. F. Selective 5-Lipoxygenase Inhibition by Zileuton in the Treatment of Relapsing Ulcerative Colitis: A Randomized Double-Blind Placebo-Controlled Multicentre Trial. Eur. J. Gastroenterol. Hepatol. 1994, 6, 209-215.
- (9) Friedman, B. S.; Bel, E. H.; Buntinx, A.; Tanaka, W.; Han, Y. H.; Shingo, S.; Spector, R.; Sterk, P. Oral Leukotriene Inhibitor (MK-886) Blocks Allergen-Induced Airway Responses. Am. Rev. Respir. Dis. 1993, 147, 839-844.
- (10) Corey, E. J.; Cashman, J. R.; Kantner, S. S.; Wright, S. W. Rationally Designed, Potent Competitive Inhibitors of Leukotriene Biosynthesis J. Am. Chem. Soc. 1984, 106, 1503-1504.
- (11) (a) Summers, J. B.; Gunn, B. P.; Martin, J. G.; Mazdiyasni, H.; Stewart, A. O.; Young, P. R.; Goetze, A. M.; Bouska, J. B.; Dyer, R. D.; Brooks, D. W.; Carter, G. W. Orally Active Hydroxamic Acid Inhibitors of Leukotriene Biosynthesis. J. Med. Chem. 1988, 31, 3-5. (b) Jackson, W. P.; Islip, P. J.; Kneen, G.; Pugh, A.; Wates, P. J. Acetohydroxamic Acids as Potent, Selective, Orally Active 5-Lipoxygenase Inhibitors. J. Med. Chem. 1988, 31, 499-500. (c) Summers, J. B.; Gunn, B. P.; Martin, J. G.; Martin, M. B.; Mazdiyasni, H.; Stewart, A. O.; Young, P. R.; Bouska, J. B.; Goetze, A. M.; Dyer, R. D.; Brooks, D. W.; Carter, G. W. Structure-Activity Analysis of a Class of Orally Active Hydroxamic Acid Inhibitors of Leukotriene Biosynthesis. J. Med. Chem. 1988, 31, 1960-1964.
- (12) (a) Tateson, J. E.; Randall, R. W.; Reynolds, C. H.; Jackson, W. P.; Bhattacherjee, P.; Salmon, J. A.; Garland, L. G. Selective Inhibition of Arachidonate 5-Lipoxygenase by Novel Acetohydroxamic Acids: Biochemical Assessment in Vitro and ex Vivo. Br. J. Pharmacol. 1988, 94, 528-539. (b) Payne, A. N.; Garland, L. G.; Lees, I. W.; Salmon, J. A. Selective Inhibition of Arachidonate 5-Lipoxygenase by Novel Acetohydroxamic Acids: Effects on Bronchial Anaphylaxis in Anaesthetized Guinea-Pigs. Br. J. Pharmacol. 1988, 94, 540-546. (c) Higgs, G. A.; Follenfant, R. L.; Garland, L. G. Selective Inhibition of Arachidonate 5-Lipoxygenase by Novel Acetohydroxamic Acids: Effects on Bronchial Anaphylaxis in Anaesthetized Guinea-Pigs. Br. J. Pharmacol. 1988, 94, 547-551.

- Satoh, Y.; Stanton, J. L.; Hutchison, A. J.; Libby, A. H.; Kowalski, T. J.; Lee, W. H.; White, D. H.; Kimble, E. F. Substituted Chromenes as Potent, Orally Active 5-Lipoxygenase Inhibitors. J. Med. Chem. 1993, 36, 3580-3594.
 Henning, R.; Lattrell, R.; Gerhards, H. J.; Leven, M. Synthesis
- (14) Henning, R.; Lattrell, R.; Gerhards, H. J.; Leven, M. Synthesis and Neuroleptic Activity of a Series of 1-[1-(Benzo-1,4-dioxan-2-ylmethyl)-4-piperidinyl]benzimidazolone Derivatives. J. Med. Chem. 1987, 30, 814-819.
- (15) Complete epoxide opening was observed when ethanol was used as the solvent. Higher pressure and/or longer reaction time resulted in significant over-reduction. See also ref 16c.
- (16) (a) Nelson, W. L.; Wennerstrom, J. E. Absolute Configuration of Glycerol Derivatives. 4. Synthesis and Pharmacological Activity of Chiral 2-Alkylaminomethylbenzodioxans, Competitive α-Adrenergic Antagonists. J. Med. Chem. 1977, 20, 880-885.
 (b) Clark, R. D.; Kurz, L. J. Synthesis of the Enantiomers of erythro-2-Oxiranyl-1,4-benzodioxan. Heterocycles 1985, 23, 2005-2012. (c) Delgado, A.; Leclerc, G.; Lobato, M. C.; Mauleon, D. Short and Enantioselective Syntheses of (R)- and (S)-2-Hydroxymethyl-1,4-benzodioxan. Tetrahedron Lett. 1988, 29, 3671-3674.
- (17) A method for the enzymatic resolution of 8-substituted 1,4benzodioxan-2-methanols, which was applied for enantiospecific synthesis of (+)- and (-)-flesinoxan was described recently; see: (a) Ennis, M. D.; Old, D. W. Enzymatic Resolution of 2-Hydroxymethyl-1,4-benzodioxans. *Tetrahedron Lett.* 1992, 33, 6283-6286. (b) Ennis, M. D.; Ghazal, N. B. The Synthesis of (+)- and (-)-Flesinoxan: Application of Enzymatic Resolution Methodology. *Tetrahedron Lett.* 1992, 33, 6287-6290.

- (18) (a) McClure, D. E.; Arison, B. H.; Baldwin, J. J. Mode of Nucleophilic Addition to Epichlorohydrin and Related Species: Chiral Aryloxymethyloxiranes. J. Am. Chem. Soc. 1979, 101, 3666. (b) Klunder, J. M.; Onami, T.; Sharpless, K. B. Arenesulfonate Derivatives of Homochiral Glycidol: Versatile Chiral Building Blocks for Organic Synthesis. J. Org. Chem. 1989, 54, 1295-1304. (c) Aigbirhio, F.; Pike, V. W.; Francotte, E.; Waters, S. L.; Banfield, B.; Jaeggi, K. A.; Drake, A. S-[1-(2,3-Diaminophenoxy)]-3'-(N-t-butylamino)-propan-2'-ol - Simplified Asymmetric Synthesis with CD and Chiral HPLC Analysis. Tetrahedron Asymm. 1992, 3, 539-554.
- (19) Optical purity was determined by integration of the signals for one of the diastereotopic C-1 methylene protons (δ 4.03 and 4.11) of the Mosher esters, which were well resolved in the 300 MHz ¹H NMR spectra recorded in benzene-d₆ but not in CDCl₃.
- (20) Summers, J. B.; Kim, K. H.; Mazdiyasni, H.; Holmes, J. H.; Ratajczyk, J. D.; Stewart, A. O.; Dyer, R. D.; Carter, G. W. Hydroxamic Acid Inhibitors of 5-Lipoxygenase: Quantitative Structure-Activity Relationships. J. Med. Chem. 1990, 33, 992– 998.
- (21) Satoh, Y.; Libby, A. H.; Powers, C.; Kowalski, T. J.; White, D. H.; Kimble, E. F. Benzoxepin and Benzothiepin Derivatives as potent, Orally Active inhibitors of 5-Lipoxygenase. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 549–552.

JM940537L